

December, 10<sup>th</sup> 2013

**Inter-laboratory Proficiency Test 1 and 2, 2013**

Dear Colleague(s),

Thank you for providing us the results of the Proficiency Tests 2013 in time.

Please find enclosed below information on the content in each ampoule from PT1 (ampoule I to V) and PT2 (ampoule VI to X).

A full report will be produced and submitted to you in February 2014.

We wish you all a Happy New Year!

Yours sincerely

Niccolò Vendramin, Anemone Ojala and Niels Jørgen Olesen

**CONTENT OF AMPOULES, PT1:**

Code	Specifications
Ampoule I:  IPNV strain Sp + VHSV DK-6137 (Hjarnø)	<p><b>IPNV strain Sp</b> Infectious Pancreatic Necrosis (IPN) virus isolated from Danish rainbow trout. Reference strain of IPNV serotype Sp.</p> <p><b>Received from:</b> National Veterinary Institute, Technical University of Denmark.</p> <p><b>Cell culture passage number:</b> 18</p> <p><b>Reference on isolate:</b> Jørgensen PEV &amp; Bregnalle F (1969) Infectious pancreatic necrosis in rainbow trout in Denmark. <i>Nordisk Veterinærmedicin</i> <b>21</b>, 142-148.</p> <p>Jørgensen PEV &amp; Grauballe PC (1971) Problems in the serological typing of IPN virus. <i>Acta Veterinaria Scandinavica</i> <b>12</b>, 145-147.</p>
	<p><b>VHSV strain DK-6137</b> The isolate originated from an outbreak of VHS with high mortality in sea water aquaculture.</p> <p><b>Received from:</b> National Veterinary Institute, Technical University of Denmark.</p> <p><b>Cell culture passage number:</b> 3</p> <p><b>Genotype:</b> la</p> <p><b>GenBank accession number:</b> <a href="#">AY546593</a></p> <p><b>Reference on isolate:</b> <a href="#">Olesen NJ, Lorenzen N &amp; Jørgensen PEV (1993). Serological differences among isolates of viral haemorrhagic septicaemia virus detected by neutralizing monoclonal and polyclonal antibodies. Diseases of Aquatic Organisms</a> <b>16</b>, 163-170.</p> <p><a href="#">Olesen NJ, Lorenzen N &amp; LaPatra S (1999). Production of neutralizing antisera against viral hemorrhagic septicaemia (VHS) virus by intravenous injections of rabbits. Journal of Aquatic Animal Health</a> <b>11</b>, 10-16.</p> <p><b>Reference on sequence and genotype:</b> <a href="#">Einer-Jensen K, Ahrens P, Forsberg R &amp; Lorenzen N (2004). Evolution of the fish rhabdovirus viral haemorrhagic septicaemia virus. Journal of General Virology</a> <b>85</b>, 1167-1179.</p>

Code	Specifications
Ampoule II: IHN virus 217/A	<p><b>IHNV 217/A</b>            First Italian IHNV isolate from rainbow trout.</p> <p><b>Received from:</b>            Received from Dr. G. Bovo, ISZ-Ve, Padova, Italy.</p> <p><b>Cell culture passage number:</b>            12</p> <p><b>Genotype:</b>            M</p> <p><b>GenBank accession numbers:</b>  <a href="#">FJ265716.1</a></p> <p><b>Reference on isolate:</b>  <a href="#">Bovo G, Giorgetti G, Jørgensen PEV and Olesen (1987). Infectious haematopoietic necrosis: first detection in Italy. Bulletin of the European Association of Fish Pathologists 7, 124.</a></p> <p><b>References on sequence and genotype:</b>  <a href="#">Johansson T, Einer-Jensen K, Batts W, Ahrens P, Björkblom C, Kurath G, Björklund H &amp; Lorenzen N (2009). Genetic and serological typing of European infectious haematopoietic necrosis virus (IHNV) isolates. Diseases of Aquatic Organisms 86, 213-221.</a></p>

Code	Specifications
Ampoule III: EHNV Isolate 86/8774	<p><b>EHNV Isolate 86/8774</b>            Australian freshwater isolate from rainbow trout from Adaminaby Trout Farm, NSW obtained in 1986 by Jeremy Langdon.</p> <p><b>Received from:</b>            Prof. Whittington, The OIE reference laboratory for EHN, University of Sidney, Australia.</p> <p><b>Cell culture passage number:</b>            9</p> <p><b>GenBank accession numbers:</b>  <a href="#">FJ433873</a>, <a href="#">AY187045</a>, <a href="#">AF157667</a></p> <p><b>Reference on isolate:</b>  <a href="#">Langdon JS, Humphrey JD &amp; Williams LM (1988). Outbreaks of an EHNV-like iridovirus in cultured rainbow trout, <i>Salmo gairdneri</i> Richardson, in Australia. Journal of Fish Diseases 11, 93-96.</a></p> <p><b>References on sequences:</b>  <a href="#">Hyatt AD, Gould AR, Zupanovic Z, Cunningham AA, Hengstberger S, Whittington RJ, Kattenbelt J &amp; Coupar BEH (2000). Comparative studies of piscine and amphibian iridoviruses. Archives of Virology 145, 301-331.</a>  <a href="#">Jancovich JK, Bremont M, Touchman JW &amp; Jacobs BL (2010). Evidence for multiple recent host species shifts among the ranaviruses (family Iridoviridae). Journal of Virology 84, 2636-2647.</a>  <a href="#">Marsh IB, Whittington RJ, O'Rourke B, Hyatt AD &amp; Chisholm O (2002) Rapid differentiation of Australian, European and American ranaviruses based on variation in major capsid protein gene sequence. Molecular and Cellular Probes 16, 137-151.</a></p>

Code	Specifications
Ampoule IV: SVCV strain 56/70	<p><b>SVCV strain 56/70</b>          Isolate from carp.          The isolate is most likely identical to the S/30 isolate described in  <a href="#">Fijan N, Petrinec Z, Sulimanovic D &amp; Zwillenberg LO (1971). Isolation of the viral causative agent from the acute form of infectious dropsy of carp. Veterinarski Archiv 41, 125-138.</a></p> <p><b>Received from:</b>          Prof. Fijan (January 1979 in a tube named Rhabdo virus carpio 56/70 and given as the reference strain of SVC virus).</p> <p><b>Cell culture passage number:</b>          Unknown.</p> <p><b>Genotype:</b>          Id</p> <p><b>GenBank accession numbers:</b>  <a href="#">Z37505.1 (Fijan)</a>, <a href="#">AJ538061.1 (S30)</a></p> <p><b>Reference on sequence (S30) and genotype:</b>  <a href="#">Stone DM, Ahne W, Denham KL, Dixon PF, Liu C-TY, Sheppard AM, Taylor GR &amp; Way K (2003). Nucleotide sequence analysis of the glycoprotein gene of putative spring viraemia of carp virus and pike fry rhabdovirus isolates reveals four genogroups. Diseases of Aquatic Organisms 53, 203-210.</a></p>
Ampoule V: EMEM	<b>Sterile medium for cell culture cultivation</b>

**CONTENT OF AMPOULES, PT2:**

Code	Specifications
Ampoule VI: ISAV Glesvaer/2/90	<p><b>ISAV Glesvaer/2/90</b></p> <p><b>Received from:</b> Dr. B. Dannevig, OIE Reference Laboratory for ISA, Oslo, Norway</p> <p><b>Cell culture passage number:</b> Unknown</p> <p><b>HPR Genotype:</b> 2</p> <p><b>GenBank accession numbers:</b> <a href="#">Y10404.1</a>, <a href="#">HQ259676</a>, <a href="#">AF283998.1</a>,</p> <p><b>References on isolate:</b> <a href="#">Dannevig BH, Falk K &amp; Namork E (1995). Isolation of the causal virus of infectious salmon anaemia (ISA) in a long-term cell line from Atlantic salmon head kidney. <i>Journal of General Virology</i> <b>76</b>, 1353–1359.</a></p> <p><a href="#">Falk K, Namork E, Rimstad E, Mjaaland S &amp; Dannevig BH (1997). Characterization of infectious salmon anemia virus, an orthomyxo-like virus isolated from Atlantic salmon (<i>Salmo salar</i> L.). <i>Journal of Virology</i> <b>71</b>, 9016-9023.</a></p> <p><b>References on sequence:</b> <a href="#">Mjaaland S, Rimstad E, Falk K &amp; Dannevig BH (1997). Genomic characterization of the virus causing infectious salmon anemia in Atlantic salmon (<i>Salmo salar</i> L.): an orthomyxo-like virus in a teleost. <i>Journal of Virology</i> <b>71</b>, 7681-7686.</a></p> <p><a href="#">Méroure E, LeBerre M, Lamoureux A, Bernard J, Brémont M &amp; Biacchesi S (2011). Completion of the full-length genome sequence of the infectious salmon anemia virus, an aquatic orthomyxovirus-like, and characterization of mAbs. <i>Journal of General Virology</i> <b>92</b>, 528-533.</a></p> <p><a href="#">Kibenge FS, Kibenge MJ, McKenna PK, Stothard P, Marshall R, Cusack RR &amp; McGeachy S (2001). Antigenic variation among isolates of infectious salmon anaemia virus correlates with genetic variation of the viral haemagglutinin gene. <i>Journal of General Virology</i> <b>82</b>, 2869-2879.</a></p> <p><b>References on genotype:</b> <a href="#">Opinion of the Panel on Animal Health and Welfare of the Norwegian Scientific Committee for Food Safety 26.01.07. Which risk factors relating to spread of Infectious Salmon Anaemia (ISA) require development of management strategies? Dok.nr.06/804, 68 pages.</a></p>

## EUROPEAN UNION REFERENCE LABORATORY FOR FISH DISEASES

Ampoule VII:  KHV-TP 30	<b>KHV-TP 30 (syn: KHV-T (for Taiwan))</b> KHV-TP 30 was isolated from koi in Taiwan and cloned for producing large plaques by Dr. Peiyu Lee, Institute of Medical Biotechnology, Central Taiwan University of Science and Technology, Dakeng, BeiTung District, TaiChung City 406, Taiwan in-2005.  <b>Received from:</b> Dr. Sven M. Bergmann, Friedrich-Loeffler-Institut (FLI), Federal Research Institute for Animal Health, Südufer 10, 17393 Greifswald-Insel Riems, Germany  <b>Cell culture passage number:</b> Unknown
Ampoule VIII:  KHV	<b>KHV 07/108b</b>  <b>Received from</b> Dr. J. Castric, ANSES, France.  <b>Reference on the isolate</b> <a href="#">Bigarre L., Baud M., Cabon J., Antychowicz J., Bergmann S.M., Engelsma M., Pozet F., Reichert M. &amp; Castric J. (2009) Differentiation between Cyprinid herpesvirus type-3 lineages using duplex PCR. Journal of Virological Methods 158, 51–57.</a>  <b>Cell culture passage number:</b> 4 in KF-1.
Ampoule IX:  Sterile pyrogenfree water	<b>Sterile pyrogen free water</b>
Ampoule X:  <i>Aphanomyces</i> <i>piscicida/</i> <i>invadans</i> spores NJM9701	<b><i>Aphanomyces piscicida/invadans</i> spores NJM9701</b>  <b>Received from:</b> Dr. Kishio Hatai, Lab Fish Diseases NVLU Tokyo, Japan  <b>Reference on isolate:</b> <a href="#">Kurata O, Kanai H &amp; Hatai K (2000). Hemagglutinating and hemolytic capacities of <i>Aphanomyces piscicida</i>. Fish Pathology - Gyobyo Kenkyu 35, 29–33.</a>